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## *Bartonella henselae* Antibodies after Cat Bite<sup>1</sup>

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**To the Editor:** *Bartonella henselae* is the causative agent of cat-scratch disease, which is the most common form of human bartonellosis (1). In immunocompromised patients, e.g., HIV-infected patients, *B. henselae* can give rise to longstanding fever, bacillary angiomatosis, and peliosis hepatitis (2). Domestic cats are the reservoir for *B. henselae*, and cat fleas transmit the organism between cats (3). The seroprevalence and culture findings of *Bartonella* spp. in cats have been shown to be low in Sweden (4,5) compared with warmer areas (6). Cat-scratch disease is most often spread from cats to humans by scratches, but other forms of transmission, including cat bites, have been suggested (7).

To determine seroprevalence of antibodies against *B. henselae* in Sweden, we used data from a recently published prospective study of patients with infected cat bites (8). In addition to the information about bites, information about cat scratches was collected by retrospective review of the patients' medical records. Serum samples were taken during the patient's first visit to a hospital and at a follow-up visit about 2 weeks later. The study was approved by the local ethics committee.

Immunoglobulin G against specific *Bartonella* spp. was detected by the immunofluorescence antibody test (1). Cell-cultivated antigens were prepared from the following strains: *B. henselae* Houston-1 (ATCC 49882), *B. henselae* Marseille (CIP 104756), *B. henselae* Berlin1 (M. Arvand), *B. henselae* K68 (E. Olsson-Engvall), *B. elizabethae* (ATCC 49927), and *B. grahamii* (ATCC 700132). The cutoff values for a positive immunofluorescence antibody test result were chosen as  $\geq 128$  for *B. henselae* and *B. grahamii* and  $\geq 256$  for *B. elizabethae*. The

titers are expressed as the reciprocal of the end-point dilution. For controls, we also analyzed serum from 117 blood donors with these antigens. The  $\chi^2$  test was used for statistical analysis.

We analyzed antibodies to *Bartonella* spp. in serum from 71 patients (51 women and 20 men), median age 47 years (range 15–85 years). Only 11 patients had fever. Cat scratches were reported for 17 patients. A single serum sample was obtained from 37 of the 71 patients, and an additional convalescent-phase sample was obtained from 34 patients after a median of 16 days (range 6–54 days).

Antibodies against any *B. henselae* strain were found for 24/71 (34%) patients, against *B. elizabethae* for 9/71 (13%), and against *B. grahamii* for 12/71 (17%). A total of 13/71 (18%) patients showed reactivity to *B. henselae* only. Antibodies to any *Bartonella* spp. were found for 28/71 (39%) of the patients. As many as 13/24 (54%) serum samples with antibodies against *B. henselae* reacted to antigens of only that species. More patients (19/71; 27%) reacted to the antigen from the cat in Sweden, K68, than to other strains. The least common reactivity found in this study was against the *B. henselae* Marseille strain.

Of the 117 controls, 1 (0.8%) had antibodies against *B. henselae* K68 antigens, 3 (2.6%) against Berlin1, 2 (1.7%) against Marseille, 1 (0.8%) against Houston-1, and 4 (3.4%) against any *Bartonella* spp. The difference between patients and controls was significant ( $p < 0.001$ ).

Seroconversion was reported for 6 of the 34 patients (18%) from which 2 serum samples were analyzed (Table). Among those who seroconverted for *B. henselae*, 1 had fever and only 2 reported having been scratched. Two of the patients who seroconverted were treated with doxycycline, and 1 was treated with ciprofloxacin. In addition, 1 patient with Sjogren disease was initially treated with penicillin, and later a hemangioma-like exanthema developed. Because of severe acne, the patient was treated with doxycycline for 6 months. The other patients who seroconverted were treated with penicillin or amoxicillin.

Seroconversion for *B. henselae* occurred in 4 patients, of which only 2 had reported a scratch. Three of these patients reacted to *B. henselae* Berlin1, and 1 reacted to the Houston-1 strain. Because symptoms did not differ between the patients who did seroconvert and those who did not, these findings could indicate subclinical infection.

The prevalence of immunoglobulin G against *B. henselae* in particular was shown to be much higher than that previously reported in Sweden (9). Earlier studies used only 2 *B. henselae* antigens (Houston-1 and Marseille) compared with the 4 different *B. henselae* antigens used in the present study. Most reactivity to *B. henselae* in the present study was directed against the Swedish isolate K68 (27%); only 0.8% of controls had antibodies against that antigen.

An increased prevalence of antibodies against *B. henselae* after exposure to cats has been reported from Spain (10). Because seroconversion against *B. henselae* occurred in 2 patients who had not been scratched, cat bites may contribute to transmission of *B. henselae*.

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Table. Titers against *Bartonella* spp. antigens in 6 patients who seroconverted

Patient sex/ age, y	Reciprocal titer		Interval, d	Antigen	High titer to other antigen
	Acute-phase serum	Convalescent-phase serum			
F/77	32	128	6	<i>B. henselae</i> Berlin1	<i>B. henselae</i> K68
F/50	32	256	17	<i>B. henselae</i> Berlin1	0
M/74	128	512	19	<i>B. grahamii</i>	<i>B. henselae</i> K68
F/62	32	128	44	<i>B. elizabethae</i>	<i>B. henselae</i> K68
M/56	32	128	15	<i>B. henselae</i> Houston-1	<i>B. grahamii</i>
F/23	32	128	10	<i>B. henselae</i> Berlin1	<i>B. grahamii</i>